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THE RING STRUCTURE IN SOME DERIVATIVES OF SORBOSE

By

Roy Lester Whistler

A Thesis Submitted to the Graduate Faculty for the Degree of

DOCTOR OF PHILOSOPHY

Major Subject: Plant Chemistry

Approved

Signature was redacted for privacy.

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HISTORY OF THE KETOSES

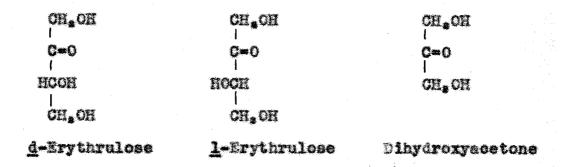
It is reasonable that in the realm of the simple saccharides the ketose sugars should hold the same high esteem as do the aldose sugars. Knowledge of the aldose sugars has been far advanced and their chemistry well established. Of the ketose sugars only one, fructose. a ketohexose, has been studied to any extent. This hiatus in the understanding of the simple saccharides can be attributed to the unavailability of the ketoses. Little by little, however, the ketoses are being studied and their characterization is now beginning to supplement aldose chemistry and to produce a more complete understanding of the simple saccharides. Such a complete understanding of monosaccharides is needful for on such knowledge rests the true structural interpretation of the complex polysaccharides, and of still higher importance, an understanding of the ultimate biologic processes.

sugars by the submergence of the carbonyl group from an end position to an internal position in the carbon chain. Generally, the carbonyl group accedes to a penultimate location in the carbon chain giving rise to the usual structures commonly ascribed to ketose sugars. Such structures will be the only ones dealt with in this dissertation.

As evidenced from the number of assymetric carbon atoms, there are possible eight ketohexoses, four ketopentoses, two ketotetroses and one ketotriose. These monosaccharides have the configurations shown below.

OH, OH	CH, OH	CH OH	CH, OH
C=0	C=0	C=0	¢=0
нсон	носн	носн	HCOH
нсон	нсон	HOCH	НОСН
нсон	нсон	НСОН	нсон
CH ₂ OH	CH, OH	CH, OH	CH, OH
<u>d</u> -Psicose	<u>d</u> -Fructose	d-Tagatose	d-Sorbose
CH OH	CH a OH	CH.OH	CH OH
¢=0	C= 0	C- 0	C=0
Носн	нсон	нсон	носн
носн	носн	нсон	нсон
носн	носн	носн	носн
CH OH	GH. OH	CH, OH	CH, OH
<u>l</u> -Psicose	1-Fructose	1-Tegatose	1-Sorbose
сн.он	CH.OH	сн.он	CH OH
¢=0	Ç=0	C=0	C=0
нсон	носн	носн	нсон
НСОН	нсон	носн	HOCH
CH, OH	CH ₂ OH	CH ₂ OH	CH 2 OH

d-Araboketose d-Xyloketose 1-Araboketose 1-Xyloketose



d-Erythrulose is not known. 1-Erythrulose (described as d-erythrulose in the older literature) has been obtained as a sirup showing a specific rotation in water of [] +11.4°.(1) It is not attacked by yeast. The ketose can be prepared through the use of Bacterium xylinum (2,3,4) or acetobacter (5) acting on a 4% solution of 1-erythitol. The sugar quickly reduces Fehling's solution. It can be precipitated with sodium bisulfite giving the addition compound. This reaction definitely shows open chain structure for the addition compound and indicates little or no tendency for the sugar to be stabilized as a lactol ring.

No ketopentoses are known in a pure crystalline form. Those recorded in the literature are probably mixtures of isomeric forms. <u>Becterium Xvlinum</u> oxidizes arabitol (6) to araboketose. <u>1</u>-Xyloketose has been found in urine during pentosuria (7-10). This sugar in water shows a specific rotation of +34.8°. It reduces Fehling's solution more rapidly than does glucose. <u>d</u>-Xyloketose

has been produced lately (11) by heating \underline{d} -xylose in dry pyridine and shows a specific rotation of -33.2° in water solution.

d-Tagatose has been prepared by the rearrangement of d-galactose through treatment with 6% calcium hydroxide (12-14). More recently d-tagatose has been prepared by the epimerization of \(\sigma \)-d-galactose (15) in dry pyridine with subsequent fermentation of the remaining galactose to alcohol. The crystalline sugar has a melting point of 162° and an optical rotation of $\sqrt{\alpha} \int_{578}^{20}$ - 3.9° in water and shows a muterotation (in 25 minutes) of +0.37° in the negative direction: thereby indicating an alpha configuration for the crystalline sugar. The diacetone derivative has been prepared (16). Methylation of tagatose (17) by Fischer's method yields &-Methyl tagatoside which has a melting point of 128° and a rotation in water of $\angle J_{578}^{70}$ + 56.8. Complete methylation gives a poor yield of pentamethyl tagatose [~720 +21.4° (in methanol) which can be hydrolyzed to a tetramethyl tagatose $\sqrt{20}_{578}$ -3.4° (in methanol). Acetylation of the sugar, employing the usual procedures of acetylation, yields a pentagetate of melting point 132° and rotation [2] \$\frac{20}{578}\$ +20.2° when dissolved in chloroform, or -25° when dissolved in methanol (18). pentaacetate is not reduced by Raney's catalyst and is

believed to be a ring form, although no proof of ring structure for tagatose or its derivatives has yet been given. <u>l</u>-Tagatose is reported as being formed by the action of alkali on <u>l</u>-sorbose (19). Recently, Glatthaar and Reichstein (20) by means of an ingenius series of reactions, have converted <u>d</u>-galacturonic acid into <u>l</u>-tagatose.

Two other presumably ketose sugars, glutose (21, 22) and galtose (23) have been recorded but are of doubtful nature. One of these may be the missing ketoallose, psicose, or perhaps one may be a ketose wherein the ketone group is on carbon atom three.

Fructose, the only ketose monosaccharide occurring in nature, will not be considered here. This sugar has been well studied. In fact, practically all of our knowledge of ketose reactivity has arisen from a study of fructose. Yet such interpretation of ketose chemistry must be held with some reserve since it is doubtful if fructose is truly representative of these sugars. Certain isolated facts arising out of the evolutionary study of other ketoses would tend to show fructose as an exception to general ketose chemistry rather than as a possessor of the representative properties of the family. With these facts in mind, it is well to proceed with a study of the history and chemistry of the ketose sorbose.

HISTORY AND CHEMISTRY OF SORBOSE

Sorbose is not commonly found in nature. produced as the result of controlled fermentation processes from the naturally occurring alcohol sorbitol. Pelouze (24, 25) in 1852 analyzed the juice of the mountain ash berries and found them to contain malic acid, calcium malate, glucose and a sugar differing in properties from those previously known. The juice of the berries was left in earthen vessels for thirteen to fourteen months. The clear supernatent liquid remaining was decanted and evaporated by gentle heat to a thick sirup. From this sirup brown crystels were deposited which were decolorized by bone black. Upon repeated recrystallization Pelouze obtained what he believed to be a pure crystalline sugar which he called sorbine (sorbose). This sugar possessed the molecular formula C.H., O. and, hence, was a monosaccharide. It could not be fermented by yeast and was not attacked by dilute acid although it turned brown and was decomposed by alkali. It rapidly reduced Fehling's solution. The optical rotation was recorded as $I = \sqrt{35.97}$. The crystals were rectangular octohedrons belonging to the right prismatic system and had a density at 15° of 1.654.

Long heating at 150-180° gave an amorphous acid substance of deep red color, which was called sorbic acid. Sorbose in fermented mountain ash berries was later observed by Byschl (26) and Berthelot (27). Delffs (28) in 1871 concluded that sorbose is not present in the original juice but is formed during the fermentation. However, since Delffs believed that sorbine belonged to the same group as mannitol, quercitol, dulcitol and persitol, it may be that he did not find sorbose but was dealing with sorbitol. Boussengault (29) found sorbitol in the freshly expressed juice, thereby demonstrating that this alcohol was not a product of fermentation. Through the use of zymitic processes, sorbose has been obtained by many workers using several bacteria but mainly Bacterium xylinum and Acetobacter (30-43). Kiliani and Schreibler (44) studied the constitution of sorbose and recognized it as a ketose. Vincent and Delachanal (45) obtained sorbitol by the reduction of sorbose with the use of sodium amalgam as the reducing agent. From this reaction it appeared possible to produce sorbose through oxidation of sorbitol.

Freund (46) also concluded sorbose to be formed by the fermentation of sorbitol in the juice of the mountain ash berry. This viewpoint has been contested by Lippman (47) who claimed that the gum over wounds in the mountain ash could be hydrolyzed to yield sorbose. These findings would suggest the natural occurrence of sorbose, but they cannot be interpreted as presenting facts antagonistic to the beliefs that sorbose may result from the fermentation of sorbitol in the juice of the mountain ash berries.

In 1904 Bertrand (48-50) published a lengthy review in which he drew several conclusions in regard to the known chemistry of sorbose.

- 1. Sorbose does not pre-exist in mountain ash berries but is formed by the oxidation of sorbitol under the influence of sorbose bacterium.
- 2. The structure of sorbose is CH_OHCHOHCHOHCHOHCOCH_OH
- 3. Since ketoses can be transformed by reduction to a mixture of two stereoisomeric alcohols of which one is identical with the original alcohol from which the ketose is derived, a method is at hand for transforming certain polyhydric alcohols into d-iditol.
- 4. The bacterium together with reduction may be employed as a means of going from an aldose to a ketose.

schlubach and Vorwerk (41) using Bertrand's method, greatly increased the yield of sorbose. They obtained between 50 and 75 per cent conversion of sorbitol. The sugar was obtained in the form of large rhombohedral crystals melting at 159-161° and had a specific rotation of $\sqrt{D} - 43.1°$ in water.

A special study of sorbose production by means of bacteria was made by Fulmer (51) and his co-workers, wherein it was found that a fifteen per cent concentration of sorbitol was optimum; the conversion to sorbose being about eighty per cent. In this work the use of Acetobacter suboxydans proved to be more expedient than Acetobacter xylinum, since the purification and separation of the sugar was more easily effected when the former bacteria were used. Employing the methods of Fulmer the U. S. Agricultural By-Products Laboratory (52) has studied the large scale production of sorbose and has successfully prepared this sugar in large quantities.

of theoretical significance is the preparation of 1-sorbose through the oxidative action of bromine water (53, 54) and of formaldehyde (55,58) on 1-sorbitol.

Dilute alkali converts 1-sorbose to 1-galactose, 1-gulose, and 1-idose (57). Reduction of the sugar by

(64) and the 2-nitrophenylosazone (65) have been prepared of 4-sorbitol and 1-1ditol (57). The phenylosezone (68. Yeast in the orystalline form. Tables have been propered (66) acid (60, 61) and is colored a purple red with an otherthe use of sodium-smalgam produces the expected mixture laevulinio aoid, humio meterial, and furfural (60, 61), the fructose-reaction with resorcinol and hydrochloric 4 1-sorbose is said to be converted to glycocollic acid does not attack the pure sugar (62). 1-sorbose gives bromine-water mixture (63). The p-bromophenylosezone gulose and 1-idose (melting point 164). Witric sold oxidation of the sugar yields xylotrihydroxyglutario 59) of 1-sorbose is identical with the osszones of acid (44). On heating with acid there is produced for the quantitative determination of 1-sorbose by treatment with onlorine and silver oxide (62). means of Febling's solution.

temperature; the rotations as compared to those above being per cent concentration and -44.8° at forty per cent concentration; the specific rotation being -42.4° at five centration in water at room temperature. They further Mutar of ation of 1-sorbose was not observed by the found that the rotation increased with increasing conobserved a decrease in specific rotation with rise in Smith and Pollons (61) esriier workers (14, 41, 67).

-39.1° at five per cent concentration and -40.6° at forty per cent concentration in water at 80°C. Recently a careful study of sorbose rotational values was undertaken by the U. S. Bureau of Standards (68). Through the use of concentrated solutions of pure sorbose in water it was found that the rotation increased slightly and then decreased so that the initial and final rotations were not widely different; the change being about 0.7° S. Mutarotation was found to be essentially complete in one hundred and twenty minutes. Exact specific rotations were found to be $\sqrt{20^{0.4} - 43.3^{\circ}}$ and $\sqrt{20^{0.4} - 43.4^{\circ}}$ at a concentration corresponding to twelve grams of sorbose in one hundred cubic centimeters of solution.

Fischer (69) using his versital method of shaking the pure sugar with a one-half to one per cent solution of dry hydrochloric acid gas dissolved in methanol. This sorboside had a melting point of 120-122° C. and a rotation of $\sqrt{20}$ - 88.7° in water. This compound was later studied by Arragon (70). It is not attacked by yeast or emulsin. Recently the stereoismeric β-methyl-1-sorboside (71) which has a melting point of 106.2° C. and a specific rotation of $\sqrt{20}$ - 39° (in water) has been prepared through the deacetylation of the β-methyl-1-sorboside tetracetate of melting point 75° C. and

rotation $\triangle J_D^{20}$ - 79.8° (in chloroform). The last compound was prepared through the action of methanol, silver carbonate and silver nitrate on \angle -acetochlorosorbose of melting point 67°C. and rotation of $\triangle J_D^{20}$ - 83.8° in chloroform. The \angle -acetochlorosorbose was prepared by treating \angle -sorbose tetraacetate (72, 73) with liquid hydrochloric acid for two hours at 0°C. in a glass bomb.

Through the direct acetylation of sorbose, Schluback and Vorwerk (41) obtained a crystalline pentagetate having a melting point of 97.5°C. and a specific rota-later obtained by Arragon (72) and was believed to be the ~-pentagoetate. However, in opposition to fructose and other sugars this compound did not form the expected halogenose. The compound further showed strong reduction to Fehling's solution. Thus it appeared possible that the acetate was a derivative of the open-chain form of That such was the case has lately been proven sorbose. by Cramer and Pacsu (74) when they were able to hydrogenate this acetate using platinum suspended in ether solution, and then, after acetylation of the products, to obtain the hexaacetates of d-sorbitol and 1-iditol. What is thought to be the true &-sorbose pentaacetate (75) has been prepared through the further acetylation of sorbose tetrascetste using acetic anhydride and two per cent sulfuric acid at 5° C. The substance showed a melting point of 95° C. and a rotation of $20 - 52.4^{\circ}$. The $20 - 52.4^{\circ}$ are sorbose pentagoetate (71) was prepared through the action of acetic anhydride and silver acetate on $20 - 40 - 40 = 113.8^{\circ}$ C. and a rotation of $20 - 74.4^{\circ}$ in chloroform.

~-Methyl-1-sorbose tetracetate of melting point 88°C. and rotation of $/\sim 7$ -31.8° has been prepared both by the methylation of \sim -sorbose tetracetate and by the acetylation of \sim -methyl-1-sorboside (70).

Using the general procedures for methylation Arragon (76) has methylated —methyl—1-sorboside to a pentamethyl derivative showing the following constants; N 1.4475,

20 19 1.108, [] - 51.5°(in methanol). This compound for the following constants; N 1.4475,

20 20 70 - 5780

4.9° (in methanol) and an %-tetramethyl sorbose [] - 5780

\$\frac{20}{-3.6°(in methanol)}\$. Methylation of the following constants; N 1.4475,

20 20 70 - 3.6°(in methanol). Methylation of the following constants; N 1.4475,

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20 20 70 - 3.6°(in methanol).

in water. Compounds of sorbose with bromal (79), resorcinol (80) and phloroglucinol (81) have been reported. Sorbose also takes up hydrocyanic acid to produce a crystalline product (44). A trinitrate of sorbose anhydride has been reported (82).

As an intermediate in the synthesis of vitamin C, Reichstein and Grüssner have prepared 2,3,4,6-diacetone sorbose (42).

d-Sorbose in the older literature was designated 1-sorbose. The d-modification has been prepared (57) through the rearrengement of d-galactose in alkali. When the products were dissolved in methanol-analine solution d-sorbose was the first to crystallize. It showed a melting point of 164°C. and a rotation of $\mathcal{L} = \mathcal{I}_{n}^{20} + 42.9^{\circ}$ (in water). The sugar has also been obtained through the rearrangement of d-gulose or d-iodose (39). Reduction of d-sorbose by sodiumamalgam produced d-iditol and d-sorbitol. The methyl-d-sorboside showed a melting point of 119°C and a rotation of $I_{\mathcal{A}_{D}}$ + 88.5° (in water). Monomethylene d-sorbose of melting point 81°C, and rotation of 27 + 25° has been produced. The phenylosazone of melting point 168°C. is identical with that of gulosazone and d-idosazone.

STATEMENT OF THE PROBLEM; DISCUSSION OF RESULTS.

With the sole exceptions of discetone sorbose and the open-chain pentascetate, the ring structures for 1-sorbose and its derivatives have remained undetermined. It has been the purpose of the work herein discussed to supply a precise structural background to this field. This has, in part, been done through theoretical considerations and through exact chemical methods.

Among the central compounds of the simple sugars are the glycosides, and any precise development of monosaccharide chemistry based upon strict constitutional assignment must in general be referred to these compounds. Thus, the assertion of their structure is of first importance. The recent development of sorbose chemistry has excluded such structural assignment, although methyl-1-sorboside (69) was one of the earliest known derivatives of sorbose. The high negative rotation (-88.7) compared with that for pure sorbose (-43.4°) indicates that this glycoside is an alpha modification (83).

Since ethylene oxide rings have never been known to form under the usual conditions for glycoside formation, and since the molecular length of a ketohexose molecule does not permit the production of a septanoid

ring, the structure of <-methyl-1-sorboside must contain either a furanoid or pyranoid ring. Wherever a sugar is found to produce glycosides containing six membered rings and also glycosides containing five membered rings, the glycosides possessing the smaller rings have shown, in every case, higher rates of hydrolysis when compared to those possessing the larger or pyranoid ring. To be strictly comparable the aglucon must be the same for the two compared rings in the same sugar, and, moreover, the glycosides must be of the same alpha or beta isomeric types. Rates of hydrolysis are then valuable in serving to distinguish glycosides containing the stable pyranoid ring from those containing the unstable furancid ring. Rates of hydrolysis, however, do not remain the same as one passes from sugar to sugar. That is to say, <-methyld-mannoside does not hydrolyze at the same rate as does ∠-methyl-d-glucoside. This is reasonable since the contribution toward reactivity by the assymmetric centers will have different values in the different sugars. No definite conclusion concerning ring structure could, therefore, be drawn unless there existed two <-methyl-1sorbosides showing among their other properties different rates of hydrolysis. The occurrence of only one ~-methyl1-sorboside excluded any exact interpretation of hydrolysis data as regarded the assignment of ring structure. The information was, however, not without some value and so the hydrolysis of the known crystalline —methyl-1-sorboside was studied.

When pure crystalline —methyl-1-sorboside was dissolved in distilled water it gave no reduction with Fehling's solution even after standing for five days. Neither was reduction toward Fehling's solution evidenced when the sorboside was allowed to stand with hundredth normal hydrochloric acid for six hours. However, a copious precipitate of cuprous oxide occurred when the glycoside was boiled one minute in tenth normal hydrochloric acid and then tested with Fehling's solution. At a temperature of 30°C. and a concentration of eight-tenths of one per cent it was found that /-methyl-1-sorboside was completely hydrolyzed in 1.75 normal hydrochloric acid in less than one day. In 0.102 normal hydrochloric acid the hydrolysis was completed in nine to ten days. In 0.017 normal acid completion of hydrolysis occurred in about thirty days. The true end point of this latter hydrolysis was indefinite due to mold growth which became noticeable after twenty-eight days. The general shape of the hydrolysis

curve for this last case is shown in Figure 1. This curve suggested that the hydrolysis was a first order reaction. The velocity constant was calculated to be 0.049, when time was taken in days. This gave a half period of fourteen and two-tenths days.

Menzies' (84) examination of methyl-d-fructofuranoside showed that this sugar exhibited perceptible reduction toward Fehling's solution after standing in distilled water for six hours. On standing six hours in one-hundredth normal hydrochloric acid, marked evidence of hydrolysis was shown when tested with Fehling's solution. At a concentration of nine-tenths of one per cent in 0.011 normal sulfuric acid and at a temperature of 20°C, methyl-d-fructofuranoside was completely hydrolyzed in less than twenty-seven days. Although complete data concerning the hydrolysis of methyl-d-fructopyranoside do not seem to have been published, yet it is known that this glycoside is quite stable toward hydrolysis even against relatively high acid concentrations.

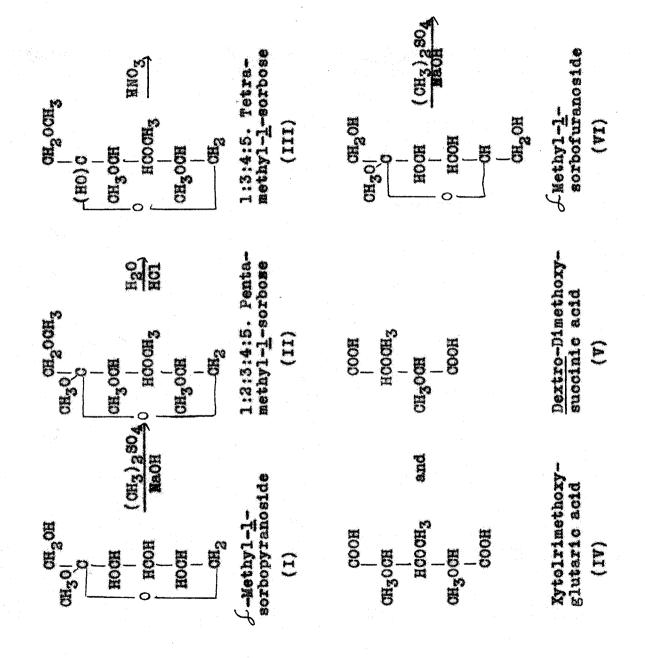
This data indicated that whereas \mathcal{S} -methyl-1-sorboside did not show the extreme stability of methyl-d-fructo-puranoside, it was evidently more stable to hydrolysis than methyl-d-fructofuranoside. Although the number of

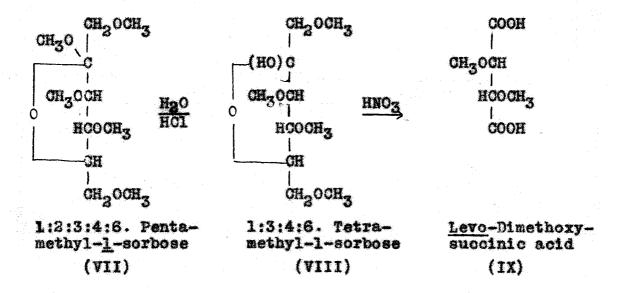
days required for the complete hydrolysis of methyl-dfructofuranoside and of \(\int \)-methyl-l-sorboside did not differ
greatly, it must be noted that the hydrolysis of the
sorboside was carried out at 30°C. in contrast to the
lower temperature of 20°C. used for methyl-d-fructofuranoside. Had the hydrolysis of \(\int \)-methyl-l-sorboside
been carried out at 20°C. the time for completion of the
reaction would have been greatly extended.

These comparisons might possibly point to \(\alpha\)-methyll-sorboside as containing the more stable or pyranoid
ring. Yet, such data cannot be relied upon because of
the previously stated reason that hydrolytic studies are
not strictly comparable between the various sugars. For
exact constitutional proof of the ring structure recourse
was made to the well established methods of oxidative
degradation. For this purpose pure crystalline \(\alpha\)-methyl\(\frac{1}{2}\)-sorboside (I or VI) was methylated to produce a liquid
pentamethyl sorbose (II or VII). This completely methylated
sugar was hydrolyzed by hot two per cent hydrochloric acid
to produce a tetramethyl derivative (III or VIII).

At the time of these preparations the pentamethyl-1-sorbose and the tetramethyl-1-sorbose were the first methylated derivatives of sorbose known. The optical rotations were taken in chloroform. A paper by Arragon (76) soon appeared announcing the preparation of these same derivatives but with rotations recorded in methanol. For comparison with Arrogon's work the optical rotations of the penta-methyl and tetramethyl sorbose were repeated in methanol. Confirmation of Arragon's rotations was obtained.

Since it is known (85) that in such a series of reactions must have contained a ring identical with that possessed the ring form remains unaltered, the tetramethyl sugar by &-methyl-1-sorboside.





The tetramethyl-1-sorbose was oxidized by concentrated nitric acid to dextro-dimethoxysuccinic acid. The latter product was obtained in good yield. No intermediate products of oxidation corresponding to those of fructose (86) were isolated. However, the presence of dextro-dimethoxy-succinic acid alone sufficed for the certain allocation of the lactol ring if consideration of an ethylene oxide structure were forgone.

If the oxygen bridge engaged carbon atom five (VIII) a large yield of <u>levo</u>-dimethoxysuccinic acid (IX) would be expected in the oxidation products of 1:3:4:6.-tetramethyl sorbose. No <u>levo</u>-dimethoxysuccinic acid could be obtained. If the oxygen bridge engaged carbon atom six (III) the oxidation products would be expected to yield xylotrimethoxyglutaric acid (IV) and <u>dextro</u>-dimethoxy-

succinic acid (v). Probably due to an insufficient amount this product, since it asserted methylation of carbon atom tained carbon atoms four and five, since no other adjacent of material no trimethoxyglutaric acid could be separated. five, alone sufficed for the elimination of the furanose carbon atoms in the tetramethyl sorbose could have given structure. Dextro-dimethoxysuccinic acid must have con-Dextro-dimethoxy succinic acid was, however, isolated in good yields. It was seen at once that the isolation of rise to an acid of this configuration.

proved definitely by the preparation of its amide and methyl These are well defined crystalline derivatives prepared by Haworth (87) as reference compounds in the sugar The presence of destro-dimethoxysucoinic acid was series.

Lethyl-1-sorboside tetrascetate, L-methyl-1-sorboside tetrataneously, with this proof of ring structure for L-methyl-1configuration for all the derivatives of this glycoside, as + 4.950) which may now be called f-methyl-1-sorbopyranoside. Simul-This evidence conclusively demonstrated a normal pyrasorbopyranoside there logically appeared the proof of ring and, hence, also for the glycoside, &-methyl-1-sorboside, for example: sorbose tetrascetate, f-ethyl-1-sorboside, noid configuration for tetramethyl sorbose (Egjas

benzoate, pentamethyl sorbose, tetramethyl sorbose, B-methylducts can now be assumed to possess a normal pyranoid ring compounds to 1-sorboside, and -chloroscetosorbose. All of these prosome of these 1-methyl-1-sorboside will be discussed later. The relationship of structure.

starting material was recovered. J-Methyl-1-sorbopyranoside stability. Neither of the above two derivatives showed retreated with several reagents designed to produce halogeno placement of the methoxyl group by chlorine when they were pyranoside the tetraacetate (76) was prepared. The glyco-In the further characterization of A-methyl-1-sorbo-These conclusions were verified by later workers sidic methyl group seemed to be extremely stable in this derivative also the glycosidic methyl group showed great In this chloride saturated with hydrochloric acid gas only the derivative for even on standing over night in acetyl tetrabenzoate was prepared for the first time. sugars.

ure 1). The close similarity in the optical changes indicated a reaction between sorbose and the ethanol solution rotary change analogous to the change occuring during the formation of &-methyl-1-sorbopyranoside in methanol (Figformation with regard to 1-sorbose it was observed that During the study of Fischer's method for glycoside solution of this sugar in ethanol underwent an optical

with formation of an ethyl-1-sorboside, which like the methyl-1-sorboside would be expected to be of the alpha configuration. The reaction was essentially complete in four hours at room temperature.

A small quantity of 1-sorbose was subjected to the action of a one per cent solution of hydrogen chloride in dry ethanol with recovery, at the end of four hours, of fine colorless needles having a melting point of 116° C, and an optical rotation of \mathbb{Z}_D^{26} - 73.9° when dissolved in water. The strong negative rotation as compared to 1-sorbose indicated an \(\frac{1}{2}\)-glycoside. On solution in dilute hydrochloric acid the hydrolysis curve followed closely that found for \(\frac{1}{2}\)-methyl-1-sorbopyranoside. Ten days were required for complete hydrolysis in tenth normal acid and approximately thirty days in 0.015 normal acid (Figure 1). The velocity constant for hydrolysis in the 0.015 normal acid was calculated to be 0.043, when time was taken in days. From this constant the half period for hydrolysis was calculated as sixteen days.

The close similarities of the rate of formation and of the rate of hydrolysis with the corresponding rates for —methyl-1-sorbopyranoside suggested that —ethyl-1-sorboside might contain a stable pyranoid ring. The truth of this assumption was proved by methods which permitted the direct chemical linkage of —ethyl-1-sorboside with

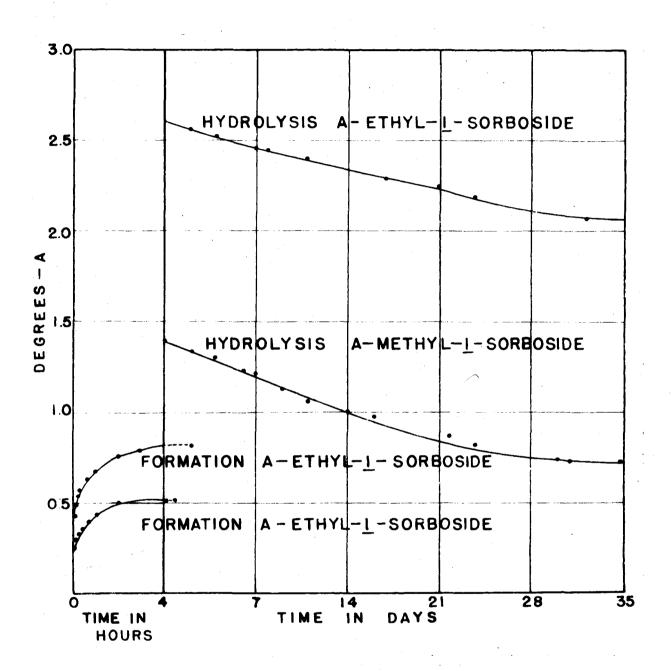


Figure 1.

S-methyl-1-sorbopyranoside.

Lethyl-1-sorboside tetraacetate which was identical with the compound obtained through the ethylation of sorbose tetraacetate. The structure of sorbose tetraacetate (72, 73) was established through the fact that acetylation of Lemethyl-1-sorbopyranoside yielded an Lethyl-1-sorbopyranoside tetraacetate (70) identical with the compound obtained through the methylation of sorbose tetraacetate. It is evident that in this series of reactions the ring structure did not change. Hence, Lethyl-1-sorboside and its tetraacetate also possess a normal pyranoid ring structure.

Through the expedient of hydrogenation and acetylation followed by subsequent isolation of the hexaacetates of d-sorbitol and 1-iditol, Cramer and Pacsu (74) have proven that the common pentaacetate of 1-sorbose is a derivative of the open chain sugar. This conclusion might have further been substantiated if it could have been shown that sorbose pentaacetate were capable of mercaptalization to yield 1-sorbose ethyl thioacetal pentaacetate. With this in mind the mercaptalization of the keto sorbose pentaacetate was undertaken. To prevent the loss of the easily hydrolysable acetyl groups, mild conditions and strictly anhydrous solutions were employed. Following the methods advanced by Wolfrom (88), for the mercaptalization of fructose, zinc

chloride was used as the catalyst. When a solution of keto sorbose pentaacetate was treated with dry ethyl mercaptan containing zinc chloride and a reaction insoluble debydrating agent, a substance was produced which did not reduce Fehling's solution. The sirup distilled in a high vacuum but with some decomposition. A specific optical rotation of $\square / 30 - 13.1^{\circ}$ in chloroform and a refractive index of N_{D}^{24} 1.5050 were found. The compound could be further demercaptalated to produce keto sorbose pentaacetate. From this evidence, it might be assumed that the compound was indeed the 1-sorbose thicacetal pentaacetate and, hence, a derivative of the open chain sugar. The material could not be obtained crystalline. Lack of time made impossible any further attempts to crystallize this compound. It is interesting, in this connection, that the non-crystalline character of the entiomorphic compound, d -sorbose ethyl thicacetal pentaacetate, has been noted by Wolfrom (89).

For the further study of ring structure in derivatives of 1-sorbose the monotosyl (mono-p-toluenesulfonyl) derivative was especially desired. The monotosylation of 1-sorbose was of interest because the reaction implied a study of the rates of reactivity for the various hydroxyl groups in the sugar. Compeditive esterification had up to this time never been studied in the ketose series. The unimolecular acylation of aldose derivatives containing both

primary and secondary hydroxyl groups led to the preferential esterification of the former (90). Among the secondary hydroxyl groups that could be present, the one adjacent to the carbonyl group, or position two in the aldose sugars, was usually the most reactive. With these considerations in mind the speculation was made that in sorbose the hydroxyl group on carbon atom one as well as the hydroxyl group on carbon atom six would show high reactivity. The hydroxyl on carbon atom three, since it is adjacent to the carbonyl group, was expected to be reactive. Since the hydroxyl group on carbon atom one was both primary and adjacent to the carbonyl group it was expected to show a very high reactivity.

Early in this work a monotrityl-1-sorbose (monotriph-enyl-methyl-1-sorbose) was prepared. This compound did not form an osazone nor give a precipitate with phenyl hydrazine. It, however, showed strong Fehling's reduction.

These facts suggested that the compound was a derivative of 1-sorbose in which carbon atom one was blocked.

When a solution of 1-sorbose in dry pyridine was treated with a molecular equivalent of tosyl chloride there was formed a compound which, though it did not react with phenyl hydrazine, showed a strong Fehling's reaction.

Again these reactions suggested that carbon atom one was blocked, while the potentially reducing carbon atom two re-

produced 1-tosyl-1-serbose, no conclusions can be drawn un-While first In the time that was had this compound could observation indicated that the monotosylation of 1-sorbose not be obtained in the pure crystalline state. til further work has been accomplished. mained open.

OPTICAL ROTATORY CONSIDERATIONS

Hudson (91) has observed a number of interesting generalizations pertinent to the optical rotations in the In his speculations on sorbose he calculated the specific rotation to be $\int (b_{sorbose} - a_{Me}) + (a_{Me} - a_{OH}) = 7$ mol. wt. = $(-17,200 + 18,500 - 8,500) \div 180 = -40$, a value quite near the observed rotation of sorbose. He believed that this result was a sufficient indication for regarding -43° as the true rotation of $\sqrt{-1}$ -sorbose, and that such a value was not an equilibrium rotation such as the value +520 for glucose. This conclusion he believed justified because the equilibrium rotations of the many rotating sugars are in all cases widely different from the rotations of their alpha and beta forms. As sorbose was later shown to be the first exception to this latter statement, it was well that Hudson limited his remarks by saying that they should be regarded as indications only, and not proofs. At this time only two crystalline ketoses, fructose and perseulose, had been observed to undergo mutarotation. Mutarotation had not been detected for tagatose, mamoketoheptose or sorbose. The reasons for this unusual lack of isomerization in solution were unexplainable, although spatial interference in ring formation was partly indicated (92).

The agree-As indicated in the historical section, the mutaropredominated when the sugar was dissolved. These theories Since that time mutarotation was observed in sorbose Pignan and Isbell tested by means of two solubility meastion of the solution was taken after equilibrium had been This logical conclusion made no claim as to what isomeric rotation observed. In the second case the specific rotaexclusively of the isomer known in the crystalline state. shaken with ice water for three minutes and the specific smallness of the mutarotation was due either to the fact found that the two specific rotations differed from each (a) that the equilibrium was established between isomers other by less than one per cent, thereby indicating that It was Pigman and Isbell believed the having only slightly different optical values, in which isomer; or (b) that the known modification greatly the equilibrium solution of sorbose was composed almost value observed, and the higher negative rotation of the ment of Hudson's calculated value for 2-1-sorbose with ease case the solution contained considerable quantities of methyl-1-sorboside were the only suggestions that the urements. In the first case an excess of surbose was The established between the sugar and the ice water. form of sorbose composed the crystalline sugar. crystalline sugar was an alpha modification. tation was very slight.

which 1-sorbose produced an open chain pentaacetate might have been claimed as evidence for postulating a free keto structure for the crystalline sugar. The extreme doubtfulness that any free sugar could exist, crystalline, in open chain form did not exclude this theory from postulation. The relative high negative rotation of sorbose compared to the very small or nil rotations of open chain sugar derivatives, however, did much to discourage any conceptions of open chain structure. The isolation of derivatives of the unknown beta form or better still the isolation of the beta form of the sugar itself would have permitted a more certain assumption of ring structure for the common crystalline sugar.

With these considerations in mind various attempts were made to prepare the beta modification. There are four general methods used for preparing the beta modifications of several of the sugars, but developed mainly for \$\beta-\frac{d}{d}\) glucose. These methods are the acetic acid method of Hudson and Dale (93), the pyridine process of Behrend (94, 95), the ammonia-alcohol method of Levene (96), and an extension of Tanret's (97-99) original procedure. Each of these methods when applied to sorbose produced no new form of this sugar.

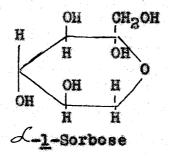
Certain salts have been known to influence the stability of one or the other isomeric forms of a dissolved sugar.

dioated that the calcium chloride addition compound did the calcium chloride addition compound. entially an open chain pentagoctate is also exhibited by obtained. dures for acetylation, the keto sorbose pentagostate was acetylating the addition compound by the general procepossess the same specific rotation as pure sorbose. the sugar component to be unaltered structurally and to resembling the rotatory changes exhibited by pure sorbose. pound showed a rapid but small upward mutarotation thus taining two molecules of water of hydration. have been stabilized in one of their alpha or beta modifithere resulted a nicely crystalline addition compound, conthat the sugare in the calcium chloride addition compounds exhibited by the pure sugar components. rotatory changes in most cases follow closely the changes orystalline double compounds with calcium chloride (100-Recently several sugars have been known to form nicely stabilize sorbose in a new form. to to ects observed initial rotation was that calculated assuming produced this acetate in good yield. All of these compounds undergo mutarotation; the When 1-sorbose was treated with calcium chloride Hence, the property of sorbose to form prefermethod (73) designed to give Such changes show Acetylation using gorbose tetrance-These facts 9

Since the very mild acetylation of the calcium chloride addition compound produced sorbose tetraacetate, known to be a derivative of sorbopyranose, added weight was given to the suggestion that crystalline sorbose was \angle -1-sorbose. At about this time also Shlubach (71) announced the preparation of a positive rotating methyl sorboside, thereby further indicating that crystalline sorbose was an alpha modification.

If it was assumed, then, that common crystalline sorbose was indeed &-1-sorbose, it followed according to Pigman's and Isbell's conclusions that a solution of sorbose in water consisted almost entirely of d-1-sorbose. This. along with the data previously mentioned, showed a strong tendency for sorbose to remain in the alpha modification. which must be the stable modification for this sugar. It was known that in water solutions the equilibrium constants (ratio of beta to alpha forms) for the different sugars varied from 0.4 to 2.3. For most of the sugars the value was 1.7 showing the predomination of the beta isomer. For certain sugars as mannose, lyxose, and rhamnose the equilibrium constant was less than one, showing that the alpha form of these compounds was predominant in the solution. Reviewing this data it appeared that in practically every case the most abundant isomer in the equilibrium solution was that in which a trans arrangement was present between the hydroxyl group on the potentially reducing

carbon and the hydroxyl on the adjacent asymmetric carbon atom. When the structure of \mathcal{L}_{-1} -sorbose was observed it was seen that such a trans arrangement existed between the hydroxyl groups.



In this respect sorbose was seen to conform to the known steric phenomena of the other sugars. Indeed, sorbose showed even greater tendency than other sugars to remain in the stable trans (alpha) form; since, according to Pigman and Isbell, almost the whole amount was retained in this form when the sugar was dissolved in water.

If Hudson's rules were assumed to hold for sorbose as for the other sugars, then the rotation of the unknown beta form could be predicted. Hudson found that the difference in molecular rotations of alpha-beta isomeric pairs was a constant value for all aldoses considered. If the same generalization were assumed to hold for the ketoses then the difference in the specific rotations between alpha and beta fructose should also be equal to the difference in specific rotation between alpha and beta sorbose. This difference for fructose was found to be

-63.3 - (-133.5) = +69.9. If now this value were added to the value -43, for \angle -1-sorbose, the value + 26.9 would be obtained as the predicted rotation for the unknown β -1-sorbose.*

At the time evidence was found for the existence of —ethyl-1-sorboside it was desired to predict the optical rotation for the substance in advance of its preparation. This prediction could easily be made through an extension of Hudson's generalizations. As mentioned above, Hudson showed that A_{Me} + A_{Me} = constant, and likewise A_{Et} + A_{Et} = constant. Without making further assumptions

A_{Et} - A_{Me} = constant, might have been expected to hold true. If this value were found to be constant then its addition to the molecular optical rotation of a methyl-glycoside would yield the molecular optical rotation of the corresponding ethyl glycoside.

When all of the available data were collected (Table I) it was seen that in reality this difference, A_{CRt} - A_{CMe}, was roughly a constant; but that the signs of the differences were not uniform. Thus, it could not be predicted whether the difference was to be added or subtracted from the molecular rotation of a methyl glycoside in order to obtain the value for the corresponding ethyl

^{*} Since sorbose was a member of an 1-series the beta sugar had to be more positive in rotational value than the alpha form. Hence, the addition of 69.9 was required.

glycoside.

Under these circumstances it was decided to take the value for fructose exclusively, for the prediction of optical rotation. The difference, A_{Et} - A_{Me}, in the case of fructose was +1,120, which indicated that the molecular rotation of the ethyl fructoside was greater than that of the methyl fructoside. If this value were added to the molecular optical rotation for <-methyl-1-sorboside the molecular optical rotation of <-ethyl-1-sorboside should have been obtained. Thus, -17,200 (molecular rotation for <-methyl-1-sorboside) + 1,120 = -16,080, the predicted molecular optical rotation for <-ethyl-1-sorboside, and -16,080 ÷ 208 = -77.3°, which is the predicted specific optical rotation for <-ethyl-1-sorboside. The true optical rotation as found was √√26 - 73.9°.

When an attempt was made to predict the optical rotation of \ll -ethyl-1-sorboside tetraacetate, the literature afforded data which, though small in extent, were consistent with themselves (Table II). In each case the ethyl glycoside tetraacetate was seen to have a numerically higher molecular rotation than the corresponding methyl compound. In the calculation with this data the average value of $A_{\rm Et}$ - $A_{\rm Me}$ could be used with impunity. If to the molecular optical rotation of \ll -methyl-1-sorboside tetraacetate, -18,820, was added the average of the differences $A_{\rm Et}$ - $A_{\rm Me}$ (-2,790)

and the molecular optical rotation thereby otained (-21,610) was divided by the molecular weight of \angle -ethyl-1-sorboside tetraacetate, its specific rotation was calculated to be -57.5. The actual value found after preparing the compound was \angle - \angle - $\sqrt{1}$ _D - 54.6°.

Table I

Rotations were taken with the D-line of sodium when the solvent was water

Substance	: Specific : Molecular: : Rotation : Rotation : Difference
L-Ethyl-d-glucoside	+150.6 +31,320
√-Methyl-d-glucoside	+158.0 +30,650 +670
8-Ethyl-d-glucoside	- 33.9 + 6,950
/3-Wethyl-d-glucoside	- 32.0 + 6,210 - 740
∠-Ethyl-d-galactoside	+ 186.8 + 38,810
∠-Methyl-d-galactoside	+ 196.6 + 38,200 + 610
P-Ethyl-d-galactoside	- 6.7 - 1,393
8-Methyl-d-galactoside	- 4.2 - 81.5 -1,310
	+ 9.95 + 2,070
∠-Methyl-l-arabinoside	+ 17.3 + 3,380 -1,290
8-Ethyl-d-fructoside	- 155.3 - 32,380
^β -Methyl-d-fructoside	-172.1 $-33,500$ $+1,120$
B-Ethyl maltoside	+ 79.2 + 29.300
8-Methyl maltoside	+ 78.8 + 28,060 +1,220
Average	(±) 994

Table II

Rotations were taken with the D-line of sodium when the solvent was chloroform

	:Specific: :Rotation:		
∠-Ethyl-d-glucoside tetraacetate ∠-Methyl-d-glucoside tetraacetate	+132 +131	+49,630 +47,420	+2,210
B-Ethyl-d-glucoside tetraacetate B-Methyl-d-glucoside tetraacetate	- 27.1 - 18.2	-10,150 - 6,520	-3,630
B-Ethyl-d-galactoside tetraacetate B-Methyl-d-galactoside tetraacetat	- 29.8 e - 25.3	-11,280 - 9,050	-2,230
P-Ethyl-d-fructoside tetraacetate P-Methyl-d-fructoside tetraacetate P-Methyl-d-fructoside tetra	-127.6 -124.4	-48,130 -45,030	-3,100
Average			2,790

EXPERIMENTAL

Preparation of 1-Sorbose

The 1-sorbose used in these researches was obtained in part as a gift from Dr. E. I. Fulmer and in part as a gift from the United States By-Products Laboratory in Ames. A further quantity of 1-sorbose was prepared according to the methods of Fulmer (51) for the bacterial oxidation of sorbitol. This sugar was purified by dissolving it in the least amount of hot distilled water required for complete solution and then adding an equal volume of 95% ethyl alcohol. The pure colorless crystals had a melting point of 164° and showed a specific optical rotation of 175-43.4°.

Preparation of &-Methyl-1-Sorboside

This substance was prepared after the method of Fischer (69). Fifty grams of powdered sorbose were placed in two liters of methanol, which had previously been dried over sodium carbonate and to which had been added 10 grams of dry hydrogen chloride. The mixture, in a gallon bottle, was shaken for two days. Then 50 grams of lead carbonate were added and the mixture shaken again for two hours. The methanol was distilled off under reduced pressure. The residual sirup was extracted repeatedly with acetone. From the acetone solution were obtained 43 grams of —methyl-1-

sorboside. This material melted at 1180.

Preparation of Tetramethyl——methyl——sorboside

In a three necked round bottom flask, equipped with a glass stirrer in the central neck and a dropping funnel in each of the two side necks, were placed 25 grams of d-methyl-1-sorboside and 15cc of water. The mixture was heated to 70°. Dimethyl sulfate was allowed to drop at the rate of one drop per second from one of the dropping funnels. Through the other funnel 60% potassium hydroxide was admitted at a rate such that the mixture in the flask was at all times slightly alkaline. In three hours 400cc of alkali and 250cc of dimethyl sulfate had been added to the reaction. The temperature was then raised to 1000 and the solution maintained at this temperature for one hour. Vigorous stirring was employed. At this time the solution was cooled and extracted with three 100cc portions of chloroform. The combined extract was washed with water and then dried over sodium sulfate. The chloroform was removed under diminished pressure. The 16 grams of remaining sirup were distilled at a pressure less than one millimeter. The liquid boiled at 90 - 950 with the bath at a temperature of 100 -1100. This sirup was remethylated according to the above

Preparation of Tetramethyl-1-sorbose

Thirty grams of tetramethyl—c-methyl-1-sorboside at 10% concentration were hydrolyzed for one and one-half hours in 2% hydrochloric acid; the temperature being maintained at 95°. On completion of the hydrolysis the solution was neutralized with lead carbonate, filtered and extracted with three 75 cc portions of chloroform. On evaporation of the chloroform under reduced pressure the remaining sirup was

a specific rotation of $\mathbb{Z}_{\mathbb{D}}^{30}$ + 4.9° in methanol. roform and M_D^{87} + 4.95° in methanol. Arragon recorded (76) showed a rotation of $D_D^{28} = \frac{-0.20 \times 100}{2 \times 4 \times 0.246} = -10.20$ in ohlolight yellow sirup of rather thick consistency. sirup distilled at a bath temperature of 125° to produce distilled at a pressure of approximately one millimeter. This sirup

Oxidation of Tetramethyl-1-sorbose

of water and the excess nitric acid removed by distilling methanol until one liter had distilled, after which the mained. perature raised carefully to 950, and the solution kept at was heated carefully to 70°. At this temperature vigorous liquid was The evaporation was further continued, replacing water by time to time and the distillation continued for six hours. at 40° under 30 millimeters pressure. reaction had ceased and a clear light yellow solution rethis temperature for two hours. the solution was replaced in the heating bath and the temcoming too violent. After the initial reaction had subsided ing bath for a few minutes to prevent the reaction from beoxidation set in and the solution was removed from the heatconcentrated nitric acid (density, 1.42). 5 grams of tetramethyl-1-sorbose were added 38.5 The mixture was poured into three times its volume taken o dryness. The product was At the end of this time Water was added from The solution esterified 00

by gently refluxing in methanol for six hours, enough nitric acid remaining to catalyze the reaction. When esterification was complete the nitric acid was neutralized by silver carbonate. The solution was evaporated to dryness and the residue extracted with ether. On evaporation of the ether a light yellow sirup remained which was distilled at about one millimeter pressure to give two fractions. The first fraction distilled at $120-125^{\circ}$ and weighed 2.1 grams; the second fraction distilled at $125-135^{\circ}$ and weighed 0.4 grams. The first fraction showed a specific rotation in chloroform of $\square \square_D^{24} = \frac{0.21 \times 100}{2 \times 4 \times 0.084} = +31.4^{\circ}.$ A micro rotation of some prepared authentic dimethyl-dextro-dimethoxysuccinic ester gave $\square \square_D^{32} + 39.1^{\circ}$ in chloroform.

Preparation of Dextro-

Dimethoxysuccinamide and <u>Dextro-</u>
Dimethoxysuccinomethylamide from Nitric Acid Oxidation Products

A 5.9 mg. sample of the light yellow sirup from the first fraction was dissolved in 3 cc of dry methanol saturated at 0° with ammonia gas. On standing in the ice box over night, long needles weighing 3.7 mg. precipitated from the solution. These crystals melted at 270° and showed specific rotation

On concentration of the alcoholic solution another milligram of orystal of dextro-dimethoxysuccinamide was obtained. = +960 in water. of $ECJ^{29}_{D} = \frac{0.15 \times 100}{(100 \ 0.940) \times 0.001504}$

3.114 mg.; N , 0.464 co.; Press., 739 mm.; M, 15.92. Found: N, 16.13. Calod. for Cell 204 N2: Temp., 370. Analysis. Sample:

A further small quantity of the first fraction was dispeared. These orystals, when reorystallized from ethyl acewater. The average yield of dextro-dimethoxysuccinomethyltate, showed a melting point of 2040 and a specific optical over night in the loe box and was then slowly concentrated. solved in 10 cc. of dry methanol which had been saturated During the evaporation burr-like clusters of crystals aprotation of $\mathbb{Z}_D^{35} = \frac{+0.80 \times 100}{(100 + 0.9706) \times 0.005967} = +130.30$ in The solution was allowed to amide in these preparations was 60%. at 00, with methyl amine.

3.763 mg.; Mg, 0.496 oc.; Press., 736 mm.; Galod. for CgH1004N2: N, 13.73 Found: N. 15.84. Temp., 350. Analysis. Sample:

The second fraction of the distillate, from the nitric quantity of unidentifiable material acid oxidation, contained some dimethyl dextro-dimethoxya small succinate and

Preparation of

of dry pyridine were added 13 cc. of acetic anhydride. After standing over night the mixture was poured into 500 cc. of ice water and extracted twice with 75 cc. portions of chloroform. The combined extract was washed successively with sodium bisulfate solution, sodium bicarbonate solution, and water. The chloroform extract was dried over sodium sulfate and the chloroform removed under diminished pressure. The sirup was crystallized from alcohol; yield 2.9 grams. The melting point was 88° and the specific optical rotation in chloroform was $\frac{2}{3x^4x^3} = \frac{-4.19x100}{3x^4x^3} = -52.4°.$ Arragon reported (70) for this compound a melting point of 88° and a specific optical rotation of $\frac{2}{3x^4} = -51.8$ in methanol.

Preparation of

 \mathcal{A} -Methyl-1-sorbopyranoside Tetrabenzoate

Eight grams of \mathcal{L} -methyl-1-sorboside were dissolved in 30 cc. of dry pyridine and to the cold solution were carefully added with stirring 24 cc. of benzoyl chloride with 60 cc. of dry chloroform. The mixture was cooled in ice for one hour and then allowed to stand at room temperature over night. The mixture was poured into 500 cc. of ice water and

the solution extracted with two 60 cc. portions of chloreform. The chloroform extracts were combined and washed with cold 5% hydrochloric acid, sodium bicarbonate solution and water. The chloroform was dried over sodium sulfate and then evaporated at reduced pressure. The remaining sirup was taken up in hot ethanol. On cooling in ice a thick light yellow sirup separated. This sirup was washed with cold ethanol and then ice water. On stirring in ice water the sirup crystallized yielding 14 grams of crystals which were not very pure. These crystals did not reduce Fehling's solution and showed an optical rotation of \mathbb{Z}_n^{28} + 6.96° in chloroform. The crystals were taken up in alcohol and the solution evaporated. After a time crystals separated. These, after being washed well with alcohol and dried, melted at 127° and showed an optical rotation of $\square \square^{27}$ = $\frac{+0.22 \times 100}{2 \times 4 \times 0.1813}$ = +15.20 in chloroform.

Analysis: Calcd. for $C_{34}H_{37}O_{9}(OCH_{3})$: OCH_{3} , 5.08. Sample: 10.044 mg., 0.960 cc. of 0.1000 N. $Na_{3}S_{2}O_{3}$. Found: OCH_{3} , 4.95.

One gram of _methyl-1-sorbopyranoside was dissolved in 50 cc. of cold acetyl chloride which had been previously

lowed to stand in the ice box over night and was then evapo-The solution was alsion of silver oxide. The benzene solution was then evapo-The orystals were taken up in benzene and the solution washed with a water suspen-Hence, the A-methyl-1-sorbopyranoside tetraacetate was recovered was 860 and the The strup was gram of which optical rotation in chloroform was [2]25 - 52.70. sirup remained From the ether solution 0.5 rated to a sirup under reduced pressure. Their melting point saturated with dry hydrogen chloride. orystallized after a few minutes. rated under reduced pressure. crystals separated. taken up in ether. unchanged.

Attempted Preparation of Acetochlorosorbose

The sirup was taken up in ether and filtered 4 grams of phosphorus pentachloride. The mixture was shaken then poured into ice water and quickly extracted with chlo-Five grams of L-methyl-1-sorbopyranoside tetraacetate this solution were added 2 grams of aluminum chloride and with occasional warming for 25 minutes. The solution was The mixture drying and evaporating the chloroform solution an amber roform and the chloroform solution washed with water. contained in a glass stoppered erlemmeyer flask. were dissolved in 20 cc. of dry chloroform. sirup remained. from norite. On evaporation 2 grams of crystals separated. These, after one recrystallization from alcohol, melted at 81° and showed an optical rotation of $\square \square_0^{25}$ - 50.5°. Hence, \square -methyl-1-sorbopyranoside tetraacetate was recovered.

Rate of Formation of -Methyl-1-sorbopyranoside

When the rate of formation of L-methyl-1-sorboside in methanol solution was studied, it was found that glycoside formation proceeded much faster than was expected.

A quantity of sorbose was shaken for several minutes with dry methanol. After some of the sugar had been dissolved 12.5 cc. of the methanol solution were placed in a 25 cc. volumetric flask. The flask was then filled to the mark with dry methanol containing 1% hydrogen chloride. The flask was shaken vigorously and a portion of the solution placed in a two decimeter polarimetric tube. The rotary changes observed are recorded in Table III. The graphic illustration of the mutarotation is shown in Figure 1. The reaction was essentially complete in four hours.

Table III

Rate of Formation of \mathcal{L} -Methyl-<u>l</u>-sorbopyranoside

Zero time taken at the point of mixing the sorbose solution with the methanol containing 1% hydrogen chloride.

Time	Observed
2 min.	0.25
4 "	0.28
6 "	0.30
6 "	0.32
8 "	0.34
10 "	0.35
15 "	0.37
80 #	0.38
87 "	0.40
55 "	0.43
90 "	0.45
2 hrs.	0.50
5.5 "	0.54
6.5 *	0.55
24 #	0.56

Rate of Hydrolysis of L-Methyl-1-sorbopyranoside

In order to test the resistance of I-methyl-1-sorboside water was prepared and allowed to stand. In six hours 1 cc. of the solution was tested with Fehling's solution. No reute, a precipitate of cuprous oxide was obtained on testing d-methyl-1-sorboside in 0.01 N acid was boiled for one min-1% solution of L-methyl-1-sorboside in 0.01 M hydrochloric a 1% solution of the pure glycoside in uodn In 24 hours a still intact, as evidenced by the lack of reaction when l no reduction. After standing five days the compound was oc. of the solution was tested with Fehling's solution. acid, likewise, showed no reduction toward Fehling's standing for 24 hours. If, however, the solution of solution duction of the Fehling's solution occured. oc. portion tested with Fehling's with Fehling's solution. toward hydrolysis, second 1

sorboside in each case was 0.8 grams per 100 cc. of solution. The concentration of -methyl-1three different acid concentrations. Hydrolysis rates were In order to make a more exact determination of the hyto be too high for accurate measurement; the hydrolysis beacid was used. The acid concentration in this case proved In the first test a solution of 1.75 normal hydrochloric drolsis rate of A-methyl-1-sorboside tests were made in followed polarimetrically.

ing completed in less than 34 hours. In the second test an acid concentration 0.102 normal in hydrochloric acid was employed. As evidenced from Table IV the hydrolysis was complete in 9-10 days. In the third test 0.015 normal acid was employed. The hydrolysis apparently required 30-31 days for completion (Table IV). The plot of this curve is shown in Figure 1. The end point was made somewhat indefinite due to mold growth which became noticeable after 28 days. Assuming the hydrolysis to be a first order reaction the velocity constant K for this last case was calculated by means of the well-known kinetic equation $K = \frac{2.303}{100} \log \frac{1}{100} = \frac{1}{100}$. The average velocity constant K was found to be 0.049 when time was taken in days. The period for half hydrolysis was then calculated to be $\frac{1}{2} = \frac{0.693}{0.049} = 14.2$ days.

Rate of Formation of \mathcal{L} -Ethyl-1-sorbopyranoside

During the study of Fischer's method (69) for glycoside formation with relation to 1-sorbose it was observed that a solution of this sugar in ethanol underwent an optical rotary change analogous to the change occurring during the formation of \sim -methyl-1-sorboside in methanol.

A quantity of ethanol which had previously been dried over sodium and distilled was saturated with pure 1-sorbose. To 12.5 cc. of this solution were added 12.5 cc. of a 1%

Table IV

Rate of Hydrolysis of -Methyl-1-sorbopyranoside
in Hydrochloric Acid

Concentration of sorboside was 0.8 grams per 100 cc. of solution. Average temperature 30°.

Time	: Acid Normality		: Acid Normal	
in days	:-2 Tube 5 :-2	Tube 6	i-L Tube 3 i-	L Tube 4
0	1.40	1.32	1.40	1.25
1	1.20	1.15	1.39	1.30
3	1.10	1.09	1.34	1.22
3	0.96	0.95	1.32	1.31
4	0.89	0.89	1.30	1.20
6	0.72	0.71	1.22	1.17
8	0.71	0.70	1.15	0.94
9	0.67	0.63	1.13	1.02
11	0.66	0.63	1.05	0.93
13	0.65		1.03	0.88
15	0.64		0.98	0.88
17	0.64		0.95	0.86
21			0.94	0.79
24			0.82	0.72
27			0.80	0.68
30			0.74	0.60
31			0.72	0.60
44			0.71	

minutes after mixing. tion of the formation of ~-methyl-1-sorboside suggested the formathese changes as compared with the changes occurring during essentially complete and illustrated graphically in Figure 1. were thoroughly mixed and the optical rotation observed two solution of hydrogen chloride an ethyl sorboside which would be of the alpha conin four hours. The mutarotation is shown in table V in dry The close similarity of ethanol. The reaction was The solutions

Rate of Hydrolysis of ~- Ethyl-1-sorbopyranoside

the hydrolysis was essentially complete in 10 days. boside a two decimeter tube. followed polarimetrically when the solution was contained in sults are recorded in acid solution was studied. corresponding methyl derivative, the rate per 100 cc. Ø order to further compare C-ethyl-1-sorboside with solution containing 0.270 grams of 0.1 normal hydrochloric acid. in Table VI where it is observed In the first case measurements were These optical changes were of d-ethy1-1-807of hydrolysis The

0.01 normal hydrochloric acid. containing 0.443 In the second case measurements were made on a solution VI indicated the reaction to be complete in about 30 grams of d-ethyl-1-sorboside per The results as recorded in 100 T

Table V

Rate of Formation of &-Ethyl-1-sorbopyranoside

Zero time taken at the point of mixing the sorbose solution with the ethanol containing hydrogen chloride.

Time in Minutes	Observed
3	0.43
3	0.44
4	0.47
5	0.49
6	0.48
8	0.51
10	0.53
12	0.54
14	0.56
17	0.56
18	0.58
21	0.60
23	0.61
29	0.63
50	0.67
87	0.72
130	0.76
180	0.78
320	0.88
2,880	0.82

Table VI

Rate of Hydrolysis of ∠-Ethyl-1-sorbopyranoside
in Hydrochloric Acid

Time in Days	:A01d _i	Normality -L	0.1:Acid	Normality -£	0.01
0		1.60		2.61	
1		1.45		2.57	
1 2		1.40		2.56	
4		1.16		2.52	
6		1.00		2.46	
7		0.93		2.45	
8		0.92		2.44	
ğ		0.98		2.43	
ıĭ				2.41	
14				2.34	
17				2.30	
ži				2.26	
24				2.20	
30				2.07	
31				2.07	
32				2.07	

days. The true end point of the hydrolysis in this case was somewhat uncertain due to mold growth which became noticeable after 26-28 days. The data for the last case is shown graphically in Figure 1. The average velocity constant K for this case was calculated to be 0.043 when time was taken in days. This gave a half period for hydrolysis of 16.1 days. It was seen at once that the rate of hydrolysis of \(\mathcal{L}\)-ethyl-1-sorboside was comparable to that for \(\mathcal{L}\)-methyl-1-sorboside.

Preparation of J-Ethyl-1-sorbopyranoside

Twenty grams of finely powdered dry sorbose were dissolved with shaking in 1500 cc. of ethanol which previously had been dried with sodium and distilled. To this solution were added 250 cc. of dry ethanol containing 20 grams of dry hydrogen chloride. The mixture at once became a light pink in color. After standing for four hours the solution was light amber in shade. A solution of 13 grams of sodium dissolved in dry ethanol was added and this mixture shaken for five minutes. Then carbon dioxide was passed through the solution for one hour. The solution was distilled under reduced pressure at a temperature no higher than 50°. The residue was extracted repeatedly with boiling ethyl acetate and ethanol. The solvents were removed from the combined extracts by distilling under reduced pressure at 50°. The

until only a brittle red gum remained. From the ethyl acetate extracts there separated on cooling a flocculent white precipitate which did not reduce Fehling's solution; yield 10 grams. This material was dissolved in hot acetone and filtered from norite. On cooling long needles separated, which had a melting point of $114-115^{\circ}$. One recrystallization from ethyl acetate raised the melting point to $115-116^{\circ}$. The specific optical rotation in water was $DD_{\rm D}^{26} = \frac{-3.40\times100}{2\times4\times0.5755} = -73.9^{\circ}$. The solution showed no mutarotation on standing for twenty-four hours.

Analysis. Calcd. for $C_6H_{11}O_5(OC_2H_5)$:

C, 46.15; H 7.69; OC_2H_5 , 21.63. Sample: 3.850 mg.; CO_2 , 6.492 mg.; H_2O , 2.664 mg. Sample: 4.808 mg.;

cc. of 0.0996 N Na₂S₂O₃, 1.40. Found: C, 45.98;

H, 7.70; OC_2H_5 , 21.77.

Preparation of $\mathcal{L}\text{-Ethyl-}\underline{l}\text{-sorbopyranoside}$ Tetraacetate

First Method

To 4 grams of L-ethyl-1-sorbopyranoside dissolved in 25 cc. of dry pyridine were added 13 cc. of acetic anlydride. After standing over night the mixture was poured into 500 cc. of ice water and extracted twice with 75-cc. portions of chloroform. The combined extract was washed successively with sodium bisulfate solution, sodium bicarbonate solution,

and water. The chloroform extract was dried over anlydrous sodium sulfate, filtered and the chloroform removed under diminished pressure. The sirup crystallized from 45% ethanol. The yield was 3 grams. The crystals had a melting point of $74-75^\circ$ and showed an optical rotation in chloroform of $\square \square_0^{26} = \frac{-3.50 \times 100}{274 \times 0.8016} = -54.6^\circ$.

Analysis. Calcd. for $C_{14}H_{19}O_{9}(OC_{2}H_{5})$:

C, 51.06; H 6.38; $OC_{2}H_{5}$, 11.96. Sample: 4.577 mg.; CO_{2} , 8.560 mg.; $H_{2}O$, 2.626 mg. Sample: 6.476 mg.;

cc. of 0.0996 N Na₂S₂O₃, 1.034. Found: C, 50.94;

H, 6.375; $OC_{2}H_{5}$, 11.95.

Second Method

Twenty grams of powdered L-sorbose tetraacetate, 75 grams of freshly prepared silver oxide and 128 grams of ethyl iodide were heated under reflux until reaction began, enough heat being generated after the reaction started to maintain the solution at the boiling point. When the reaction subsided the solution was boiled for two and one-half hours longer under reflux. At this time the ethyl iodide was distilled off and the residue extracted with ether. The ether was removed under diminished pressure and the residue taken up in hot petroleum ether (B.P. 30-49°). From this solution long needles crystallized. The yield was 14 grams. Recrystallization from 45% ethanol produced

fine, soft needles having a melting point of 74° and a specific optical rotation of $\mathbb{Z} \mathbb{J}_D^{26}$ - 54.6° in chloroform. No lowering in melting point was shown when some of these crystals were mixed with crystals obtained by the first method.

Study of the Crystallization of 1-Sorbose

In an attempt to isolate \mathcal{L} -1-sorbose the crystallization of sorbose from various solvents was studied.

Ten grams of sorbose were dissolved in 12 cc. of warm concentrated ammonium hydroxide. When complete solution of the sugar was attained 100 cc. of absolute ethanol were added. In a short time crystals appeared. The solution was then filtered by suction and the crystals washed with absolute alcohol and dried. The yield was 6 grams. The optical rotation of $\square \square_D^{33} - 42.5^\circ$ (c, 2 in water) showed the recovery of \square -1-sorbose.

To 5 grams of sorbose dissolved in 15 cc. of hot pyridine were added 55 cc. of absolute alcohol. In four hours the precipitated crystals were filtered, washed with absolute alcohol and dried; yield 2.5 grams. Rotation $\int_{\mathbb{R}} \int_{\mathbb{R}}^{33} - 43.1^{\circ}$ (c, 4 in water).

To 5 grams of sorbose dissolved in 5 cc. of water and cooled to 0° were added 10 cc. of glacial acetic acid. On standing a short time crystallization occured. The crystals

after washing with absolute ethanol and drying weighed 3 grams. The rotation was $\square \square_D^{33}$ - 43.4 (c, 2 in water).

A solution of 5 grams of sorbose dissolved in 5 sc. of hot water was heated to 100° c. Then 10 cc. of glacial acetic acid which had been heated to 100° were added with vigorous stirring to the sorbose sirup. On cooling the solution deposited 3 grams of crystals having an optical rotation of $\square \square_{D}^{33}$ - 43.4 (c, 2 in water).

Preparation of a Calcium Chloride Compound of -1-Sorbose

Twenty grams of 1-sorbose were dissolved in 25 cc. of warm water and 18 grams of calcium chloride slowly added with stirring. After effecting complete solution by stirring and heating on a hot-plate, the solution was placed in a desiccator over phosphorus pentoxide. In three weeks the solution had turned to a thick mush of crystals. were stirred with absolute ethanol, filtered and washed free from sirup with absolute ethanol. The yield of dry crystals was 15 grams. The melting point was 157° or 159° corrected. The substance melted with decomposition. Recrystallization by dissolving in water and slow concentration of the sirup in a desiccator produced fine crystals melting at 159° (corrected). The specific optical rotation in water two minutes after solution was $\sqrt{27}_D^{29} = \frac{-1.80 \times 100}{2 \times 4 \times 0.9293} = -24.2^{\circ}$. The compound underwent a small upward motarotation of about 0.3°. The mutarotation was complete in 15 minutes producing a final rotation of $\sqrt{29}$ - 23.9° as shown in Table VII.

If it were assumed that the sugar component in the compound $C_0H_{1,2}O_0$. $CaCl_2$. $2H_2O$ was unaltered structurally and possessed the same specific rotation as pure sorbose, then the value \sim could have been caldulated. Making this assumption the value of \sim as -2.50° was calculated for 1.3029 grams

Table VII

Mutarotation of Sorbose-Calcium Chloride

Addition Compound in Water

Run	Initial LL729	Final 29 EST 1	Change
1	- 24.15°	- 23.80	0.30
2	- 24.2 0	- 23.90	0.30
3	- 24.2 0	- 23.80	0.40
4	- 24.3 0	- 34.010	0.30
Mean	- 34.2 0	- 23.90	0.30

of the compound, containing 0.719 grams of sorbose. This value was in good agreement with the observed value of which was 2.520.

Analysis. Calcd. for $C_6H_{12}O_6 \cdot CaCl_2 \cdot 2H_2O$: Ca, 12.25; C1, 21.72. Sample: 0.2229 g.; 16.75 cc. of 0.08 N KMnO₄. Sample: 0.2100 g.; 0.1844 g. of AgCl. Found: Ca, 12.10; Cl, 21.74.

A second and faster method was found for the preparation of the calcium chloride addition compound of 1-sorbose. In this method a sirup was prepared by dissolving 20 grams of sorbose in 25 c. of hot water and adding 18 grams of calcium chloride. The sirup was then diluted with three times its volume of absolute ethanol and the mixture placed in the ice box. On the addition of ether in small quantities a good precipitation of the addition compound soon appeared. The yield was 8 grams. The first crop of this material showed a melting point of 159° (corrected). No change in melting point was observed on mixing this substance with the addition compound prepared by the first method. The optical rotation was $\mathbb{Z}_{0}^{27} = \frac{-2.30 \times 100}{2 \times 4 \times 1.188} = -24.2°$.

Acetylation of the Calcium Chloride Addition Compound of \mathcal{L} -1-Sorbose

First Method

To 4 grams of \mathcal{L} -1-sorbose-calcium chloride addition compound dissolved in 80 cc. of pyridine were added 60 cc. of acetic anhydride. The mixture was cooled in ice and then placed in the ice box for two days. At the end of this time the mixture was poured, with stirring, into one liter of ice water and extracted three times with 75 cc. portions of chloroform. The combined extract was washed successively with 5% hydrochloric acid, sodium bicarbonate solution, and water. The chloroform extract was dried over anhydrous sodium sulfate and the chloroform removed under diminished pressure. The sirup was crystallized from ethanol. The yield was I gram. The melting point was 99° and no depression of melting point occured when the compound was mixed with authentic keto sorbose pentaacetate. Thus, this type of acetylation led to an open chain derivative.

Second Method

Four grams of the sorbose-calcium chloride double compound were dissolved in 150 cc. of acetic anhydride containing 1.5 grams of freshly fused zinc chloride. After cooling in ice this solution was placed in the ice box for two days. The solution was then poured into a liter of ice water and extracted with three 75 cc. portions of chloroform. The combined extracts were washed with sodium bicarbonate and then water. After drying over sodium sulfate the chloroform was removed under reduced pressure. The remaining sirup crystallized from ethanol yielding about one gram of crystals. The melting point was 98-99° and showed no depression when the crystals were mixed with crystals of authentic keto sorbose pentaacetate. Thus, this method of acetylation also led to the formation of an open chain derivative.

Third Method

To 3 grams of the L-1-sorbose-calcium chloride addition compound dissolved in 30 cc. of cold pyridine there were added at intervals and with constant stirring a total of 30 cc. of acetic anlydride. The mixture was stirred in an ice bath for four hours and then poured into a liter of water and extracted with three 75 cc. portions of chloroform. The chloroform extracts were combined and washed with sodium bicarbonate solution and water. After drying over sodium sulfate the chloroform was removed under diminished pressure. The sirup was taken up in ethanol. On standing 0.036 grams of crystals appeared. Combined with authentic keto sorbose pentaacetate some of these crystals showed a mixed melting point of 97°. The specific rotation was Ler_{D}^{25} +1.6.

Hence, this material was <u>keto</u> sorbose pentaacetate. The remaining sirup was taken up in chloroform and filtered from norite. On the addition of ether and petroleum ether (BP 30-40°) 1.404 grams of crystals were obtained. Their melting point was 97°. Mixed with authentic <u>keto</u> sorbose pentaacetate a melting point of 80° was found. When the crystals were mixed with authentic \mathcal{L} -1-sorbose tetraacetate the melting point of the mixture was 97°. The specific optical rotation was $\square \square^{27} - 19 \cdot 1°$. Hence, by this process of acetylation a good yield of \mathcal{L} -1-sorbose tetraacetate was obtained.

Preparation of 1-Sorbose Thioacetal Pentaacetate

In 60 cc. of ethyl mercaptan dried over drierite were dissolved 8 grams of freshly fused zinc chloride. To this solution were added 15 grams of fresh drierite and 15 grams of anhydrous sodium sulfate. Then 13 grams of dry keto sorbose pentaacetate were added and the solution cooled in ice. The mixture was allowed to stand in the ice box for one day. The solution was poured into 200 cc. of a saturated solution of sodium bicarbonate and the precipitate filtered off. The filtrate was extracted with three 50 cc. portions of chloroform. The precipitate was removed to a beaker and extracted twice with 500 cc. portions of chloroform. The combined chloroform extracts were dried over

sodium sulfate, filtered, and the chloroform removed under diminished pressure. To the 13 grams of residual sirup petroleum ether (B.P. 30-40°) was repeatedly added and distilled to remove all traces of chloroform. The light yellow viscous sirup could not be made to crystallize. It showed no reduction to Fehling's solution. The refractive index was N_D^{24} 1.5030 and the specific optical rotation was $D_D^{27} = \frac{-3.53 \times 100}{2 \times 4 \times 3.612} = -12.2°$ in chloroform. Mutarotation was not observed. The sirup distilled in a pressure of less than one millimeter and at a temperature of 200°. The distillate was colored a light yellow evidencing some decomposition. After dissolving in ether and filtering from norite the almost colorless distillate showed a refractive index of N_D^{24} 1.5050 and a specific optical rotation of $D_D^{30} = \frac{-0.35 \times 100}{2 \times 4 \times 0.3323} = -13.15°$.

Analysis. Caled. for (C2H5S)2 C16H22O10: S, 12.98. Sample: 0.00410 g.; 2.34 cc. of 0.015 N NaOH. Found: S, 13.7.

Demercaptalization

Five grams of the above mercaptalated sirup were dissolved in 20 cc. of acetone and 10 g. of cadmium carbonate were added. The mixture was stirred in a three necked flask by a strongly driven glass stirrer. Then 10 grams of mercuric chloride dissolved in 14 cc. of acetone were slowly added. While the mixture was continually stirred small quantities of ca'mium carbonate were added from time to time. After one and a half hours the temperature of the solution was raised to 40° for fifteen minutes. Then the mixture was filtered and the precipitate washed with acetone. The acetone filtrates were combined. After the addition of fresh cadmium carbonate, the acetone was removed under diminished pressure. The dry residue was extracted with chloroform and the chloroform extract evaporated under reduced pressure. On the addition of ether to the residue, white crystals melting at 98° were obtained. On mixing some of these crystals with crystals of authentic keto sorbose pentaacetete the melting point remained at 98°.

Preparation of Monetrity1-1-sorbose

To 5 grams of sorbose dissolved in 30 cc. of dry pyridine were added 8 grams of trityl chloride. The mixture was allowed to stand at room temperature over night. It was then poured into a liter of ice water. After decanting the water from the precipitated sirup, the sirup was dissolved in 75 cc. of chloroform and the chloroform solution washed with 5% hydrochloric acid, sodium bicarbonate and water. The chloroform solution was dried over sodium sulfate and filtered from norite. On adding alcohol and evaporating

The crystals gave no orystals melted at 87º and showed an optical rotation precipitate when treated with phenyl hydrazine. 6 grams of crystals were obtained. 21.70 in chloroform.

Preparation of Monotosyl-1-sorbose

The crystalline material did not form an osazone when treated was dissolved in hot ethyl acetate and filtered from norite. with phenyl hydrazine. The crystals did not seem to be comchloride. After filtering and cooling the solution deposi-5 grams of 1-sorbose dissolved in 100 cc. of cold, ted 2 grams of crystals having a melting point of 1400 and Then an equal volume of petroleum ether (B.P. 30-400) was Lead carbonate was added to remove any remaining hydrogen The strupy bottom layer Lack of time prevented the purification of mixture was allowed to stand over night in the ice box. dry pyridine were added 5.2 grams of tosyl chloride. $=\frac{-.60x100}{4x4x0.2095}$ an optical rotation in water of [2737 added and the top layer removed. this substance. pletely pure.

SUMMARY

- 1. —Methyl-1-sorboside was completely methylated to a pentamethyl derivative, which on hydrolysis of the glycosidic methyl group yielded a tetramethyl-1-sorbose (CD²⁸ + 4.95°). The oxidation of this latter compound by means of nitric acid resulted in the production of dextrodimethoxysuccinic acid. The isolation of this acid was taken as sufficient evidence for the assumption of a normal pyranose structure in the tetramethyl-1-sorbose and, hence, also in the parent compound —methyl-1-sorboside and its derivatives.
- 2. \(-\text{Methyl-1}\)-sorbopyranoside tetrabenzoate was prepared.
- 3. \(\angle \text{Ethyl-1} \text{sorbopyranoside} \) and \(\angle \text{ethyl-1} \text{sorbopyran-} \) oside tetreacetate were prepared and characterized. By means of a series of well established reactions these compounds were linked definitely to \(\angle \text{methyl-1} \text{sorbopyranoside}, \) thus proving the presence of a pyranoid ring in their structures.
- 4. The rates of formation and the rates of hydrolysis for ~methyl-1-sorbopyranoside and √-ethyl-1-sorbopyranoside were determined.

- 5. A calcium chloride addition compound of \mathcal{L} -l-sorbose was prepared and characterized.
- 6. A sirupy 1-sorbose thioacetal pentaacetate was prepared from keto sorbose pentaacetate.
- 7. A monotrityl and a monotosyl derivative of 1-sorbose were prepared.

BIBLIOGRAPHY

- 1. Müller, Montigel and Reichstein, Helv. Chim. Acta., 20, 1468 (1937)
- 2. Bertrand, Bull. soc. chim. biol., 23, 681 (1904)
- 3. Bertrand, Compt. rend., 130, 1330 (1900)
- 4. Bertrand, Ann., 3, 259 (1904)
- 5. Bertrand, Chem. Weekblad, 10, 718 (1913)
- 6. Bertrand, Bull. soc. chim., 19, 259 and 347 (1898)
- 7. Levene and LaForge, J. Biol. Chem. 18, 319 (1914)
- 8. Greenwald, ibid., 88, 1 (1930)
- 9. Greenwald, ibid., 89, 501 (1930)
- 10. Hiller, ibid., 30, 129 (1917)
- 11. Schmidt and Treiber, Ber., 66, 1765 (1933)
- 12. de Bruyn and van Ekenstein, Rec. trav. chim., 16, 262 (1897)
- 13. de Bruyn and van Ekenstein, ibid., 18, 72 (1899)
- 14. de Bruyn and van Ekenstein, ibid., 19, 5 (1900)
- 15. Danilowa and Schantarowisch, Ber., 63, 2269 (1930)
- 16. Reichstein and Bosshard, Helv. Chim. Acta., 17, 753 (1934)
- 17. Khowvine and Tomoda, Compt. rend., 205, 736 (1937)
- 18. Khowvine and Tomoda, ibid., 205, 1414 (1937)
- 19. de Bruyn and van Ekenstein, Rec. trav. chim., 19, 11 (1900)
- 20. Glatthaar and Reichstein, Helv. Chim. Acta., 20, 1537 (1937)

- 274 ohim., 16, trav. de Bruyn and van Ekenstein, Rec. (1897) 31.
- 73 (1899) de Bruyn and van Ekenstein, 1bid., 18,
- 262 and 269 (1897) 16, de Bruyn and van Ekenstein, ibid.,
- (1853)Pelouze, Ann. chim. phys., (3) 35, 322 34.
- 25. Pelouze, Ann., 83, 47 (1852)
- d Pharm., 78, 188 (1854) Archiv. Byschl, 36.
- (1857)chim. phys., (3) 50, 323 Berthelot, Ann.
- 28. Delffs, J. Chem. Soc., 24, 1043 (1871)
- Boussengalt, Compt. rend., 74, 939 (1872) 98
- 50. Matrot, 1bid., 125, 874 (1897)
- Bertrand, Bull. soc. chim., (3) 19,502 (1898)
- Bertrand, Ann. Inst. Past., 12, 385 (1898) 33
- Bertrand, Bull. soc. chim., (3) 15, 27 (1896) 33
- Bertrand, J. prakt. Chem., (2) 43, 545 (1890)
- Compt. rend., 127, 728 (1898) Bertrand,
- 36. Bertrand, 1bid., 126, 762 (1898)
- Wincent and Meunier, ibid., 127, 760 (1898) 37.
- 38. Emmerling, Ber., 32, 541 (1899)
- van Ekenstein and Blanksma, Rec. trav. ohim. 27, 1 (1908) 39.
- Kluyver and de Leew, Tijdschr Verg-Geneesh., 10, 170 (1934) \$
- Schlubach and Vorweck, Ber., 66, 1251 (1933) 41.
- Relohstein and Gritssner, Helv. Chim. Acta., 17, (1934) 423.
- 2 225 Micheel, Kraft and Lohman, Z. physiol. Chim., (1934) 43.

- 44. Kiliani and Schreibler, Ber., 21, 3276 (1888)
- 45. Vincent and Delachanal, Compt. rend., 125, 716 (1897)
- 46. Freund, Monatsh., 11, 560 (1890)
- 47. Lippman, Ber., 53, 2069 (1920)
- 48. Bertrand, Compt. rend. 139, 802 (1904)
- 49. Bertrand, ibid., 139, 985 (1904)
- 50. Bertrand, Ann. chim. phys. (8) 3, 181 (1904)
- 51. Fulmer, Dunning, Guymon and Unterkophr, J. Am. Chem. Soc. 58, 1012 (1936)
- 52. Wells, Stubbs, Lockwood and Roe, J. Ind. Eng. Chem., 29, 1385 (1937)
- 53. Talen, Rec. trav. chim., 44, 891 (1925)
- 54. Votocek and Lukes, ibid., 44, 345 (1925)
- 55. Küster and Schoder, Z. physiol. Chem., 141, 110 (1924)
- 56. Inghilleri, ibid., 71, 105 (1911)
- 57. de Bruyn and van Ekenstein, Rec. trav. chim., 19, 1 (1900)
- 58. Fischer, Ber., 20, 827 (1887)
- 59. Maquenne, Compt. rend., 112, 799 (1891)
- 60. Wehmer and Tollens, Ann., 243, 320 (1888)
- 61. Smith and Tollens, Ber., 33, 1285 (1900)
- 62. Fischer and Thierfelder, ibid., 27, 2031 (1894)
- 63. Fenton and Gostling, J. Chem. Soc., 73, 557 (1897)
- 64. Neuberg and Heymann, Chem. Zentral., 1 (1902) 861, 1078, 1241
- 65. Reclaire, Ber., 42, 1424 (1909)
- 66. Smith and Tollens, Z. Ver. deut. Zucker-Ind., 37, 525, (1900)

- 67. Rüber, Saertrykk av. Tidsskrift for Kjemi ag. bergvesen, m 10, p. 16 (1932)
- 68. Pigman and Isbell, Bur. Standards J. Research, 19, 443 (1937)
- 69. Fischer, Ber., 28, 1145 (1895)
- 70. Arragon, Bull. soc. chim. biol., 17, 831 (1935)
- 71. Schlubach and Graefe, Ann., 532, 211 (1937)
- 72. Arragon, Compt. rend., 196, 1733 (1933)
- 73. Arragon, 1bid., 198, 1508 (1934)
- 74. Cramer and Pacsu, J. Am. Chem. Soc., 59, 1467 (1937)
- 75. Arragon, Compt. rend., 205, 735 (1937)
- 76. Arragon, Bull. soc. chim. biol., 18, 1336 (1936)
- 77. Bosshard and Reichstein, Helv. chim. Acta., 18, 959 (1935)
- 78. de Bruyn and van Ekenstein, Rec. travl. chim., 22, 159 (1903)
- 79. Hanriot, Bull. soc. shim., (4) 5, 819 (1909)
- 80. Fischer and Jennings, Ber., 27, 1355 (1894)
- 81. Councier, Chem. Z., 20., 586 (1896)
- 82. Will and Lenze, Ber., 31, 79 (1898)
- 83. Hudson and Brauns, J. Am. Chem. Soc., 38, 1216 (1916)
- 84. Menzies, J. Chem. Soc., 2238 (1932)
- 85. Haworth, "Constitution of Sugars", Edward Arnold & Co., London (1929)
- 86. Haworth, Hirst and Learner, J. Chem. Soc., 1040 (1927)
- 87. Haworth and Jones, ibid., 2349 (1927)
- 88. Wolfrom and Thompson, J. Am. Chem. Soc., 56, 880 (1934)

- 89. Wolfrom, Private Communication
- 90. Lieser and Schweizer, Ann., 519, 271 (1935)
- 91. Hudson, "Relation Between Rotatory Power and Structure in the Sugar Group", Sci. Paper, U. S. Bur. Standards, No. 533 (1926)
- 92. Levene, J. Biol. Chem., 59, 135 (1924)
- 93. Hudson and Dale, J. Am. Chem. Soc., 39, 320 (1917)
- 94. Behrend, Ann., 353, 106 (1907)
- 95. Behrend, ibid., 377, 320 (1910)
- 96. Levene, J. Biol. Chem., 57, 329 (1923)
- 97. Tanret, Bull. soc. chim., 13, 728 (1895)
- 98. Tanret, ibid., 15, 349 (1896)
- 99. Whistler and Buchanan, Unpublished Manuscript
- 100. Dale, J. Am. Chem. Soc., 51, 2788 (1929)
- 101. Isbell, Bur. Standards J. Research, 5, 741 (1930)
- 102. Dale, J. Am. Chem. Soc., 56, 932 (1934)
- 103. Austin and Walsh, ibid., 56, 934 (1934)
- 104. Hann and Hudson, ibid., 50, 2075 (1937)